Coordination of different ligands to copper(II) and cobalt(III) metal centers enhances Zika virus and dengue virus loads in both arthropod cells and human keratinocytes


**A R T I C L E   I N F O**

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- Zika virus
- Dengue virus
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- Thiosemicarbazones
- 1,10-phenanthroline

**A B S T R A C T**

Trace elements such as copper and cobalt have been associated with virus-host interactions. However, studies to show the effect of conjugation of copper(II) or cobalt(III) metal centers to thiosemicarbazone ligand(s) derived from either food additives or mosquito repellent such as 2-acetylethiazole or citral, respectively, on Zika virus (ZIKV) or dengue virus (serotype 2; DENV2) infections have not been explored. In this study, we show that four compounds comprising of thiosemicarbazone ligand derived from 2-acetylethiazole viz., (E)-N-ethyl-2-[1-(thiazol-2-yl)ethylidene]hydrazinecarbothioamide (acetylethTSC) (compound 1), a copper(II) complex with acetylethTSC as a ligand (compound 2), a thiosemicarbazone ligand-derived from citral (compound 3) and a cobalt(III) complex with a citral-thiosemicarbazone ligand (compound 4) increased DENV2 and ZIKV replication in both mosquito C6/36 cells and human keratinocytes (HaCaT cells). Treatment of both cell lines with compounds 2 or 4 showed increased dengue viral titers at all three tested doses. Enhanced dengue viral plaque formation was also noted at the tested dose of 100 μM, suggesting higher production of infectious viral particles. Treatment with the compounds 2 or 4 enhanced ZIKV and DENV2 RNA levels in HeLa cell line and primary cultures of mouse bone marrow derived dendritic cells. Also, pre- or post treatments with conjugated compounds 2 or 4 showed higher loads of ZIKV or DENV2 envelope (E) protein in HaCaT cells. No changes in loads of E-protein were found in ZIKV-infected C6/36 cells, when compounds were treated after infection. In addition, we tested bis(1,10-phenanthroline)copper(II) chloride ([Cu(phen)2]Cl2, (compound 5)) and tris(1,10-phenanthroline)cobalt(III) chloride ([Co(phen)3]Cl3, (compound 6)) that also showed enhanced DENV2 loads. Also, we found that copper(II) chloride dehydrate (CuCl2·2H2O) or cobalt(II) chloride hexahydrate (CoCl2·6H2O) alone had no effects as "free" cations. Taken together, these findings suggest that use of Cu(II) or Co(III) conjugation to organic compounds, in insect repellents and/or food additives could enhance DENV2/ZIKV loads in human cells and perhaps induce pathogenesis in infected individuals or individuals pre-exposed to such conjugated complexes.

**Importance:** Mosquito-borne diseases are of great concern to the mankind. Use of chemicals/repellents against mosquito bites and transmission of microbes has been the topic of interest for many years. Here, we show that thiosemicarbazone ligand(s) derived from 2-acetylthiazole or citral or 1,10-phenanthroline upon conjugation with copper(II) or cobalt(III) metal centers enhances dengue virus (serotype 2; DENV2) and/or Zika virus (ZIKV) infections in mosquito, mouse and human cells. Enhanced ZIKV/DENV2 capsid mRNA or envelope protein loads were evident in mosquito cells and human keratinocytes, when treated with compounds before/after infections. Also, treatment with copper(II) or cobalt(III) conjugated compounds increased viral titers and number of plaque formations. These studies suggest that conjugation of compounds in repellents/essential oils/natural products/
food additives with copper(II) or cobalt(III) metal centers may not be safe, especially in tropical and subtropical places, where several dengue infection cases and deaths are reported annually or in places with increased ZIKV caused microcephaly.

1. Introduction

Insect bites, food or crop damage and transmission of pathogens to humans, animals and plants has led to the extensive use of insecticides, pesticides and insect attractants or repellents [1,2]. Use of these substances has substantially increased in military personnel and in agriculture or horticulture practices. Over a decade, several of these chemicals have been used extensively in all parts of the world. Some synthetic compounds or chemicals are of major concern to the environment and human health and they have been withdrawn due to the adverse effects and serious human health hazards [3–8]. One of the most recent study tested that several of the repellents including citronella candles had no effects and did not reduced mosquito attraction to humans [9]. The disadvantages of some insecticides have promoted the use of plant-based essential oils and their derived natural products, belonging to several herb species that posses repellent activities [7,10]. Some of the essential oils, herbal skin products and non-natural inorganic insecticides (the contact insecticides that are toxic to insects upon direct contact) have been shown to contain metals such as arsenates, copper, cobalt and fluorine compounds in small amounts and sulfur as a major component [11]. Also, both copper sulfate and cobalt chloride has been shown to improve the varieties in rice and vegetables that are mostly consumed in Australia and some parts of Asia [12]. In addition, complex mixtures of volatile organic compounds produced from plant-based essential oils or natural products such as citral have been extensively used as repellents to prevent transmission of human pathogens through biting arthropods such as mosquitoes [7,10,13].

Mosquitoes are the primary vectors responsible for the transmission of arthropod-borne flaviviruses that cause microcephaly (Zika Virus), dengue hemorrhagic fever, yellow fever, West Nile neuroinvasive disease (WNND) and Japanese encephalitis. With regard to the global impact from arthropod-borne diseases, dengue is the most important human pathogen (that exist as four serotypes; DENV-1, 2, 3 and 4) that should be addressed promptly. According to the 2009 WHO criteria, dengue cases have been classified according to the levels of severity. Dengue infections without warning signs or with warning signs (such as abdominal pain, persistent vomiting, fluid accumulation, mucosal bleeding, liver enlargement, increasing hematocrit with decreased platelet numbers and lethargy) were considered as less severe. However, dengue infections with severe plasma leakage, bleeding or organ failures were considered as severe cases [14,15]. Approximately 20,000–25,000 deaths and 50–100 million dengue infections with 500,000 hospitalizations are reported, annually [16]. Also, there are no fully effective vaccines available for human use and prevention of dengue [17,18]. Nevertheless, recently, WHO immunization group (Strategic Advisory Group of Experts; SAGE) has recommended the use of partially effective dengue vaccine (a live attenuated tetravalent dengue vaccine developed by Sanofi Pasteur; CYD-TDV, named Dengvaxia) that has been licensed and used in 11 countries including Brazil, Mexico, Singapore, Thailand and Indonesia [19]. Related to dengue, Zika virus (ZIKV) is an emerging infectious agent transmitted and spread by the daytime active Aedes mosquitoes, such as A. aegypti and A. albopictus. ZIKV can be transmitted by blood transfusions, sexual and placental leakage from the mother to the fetus [20,21]. An important public health concern has been declared due to global epidemic of ZIKV infections. The association between ZIKV infection during pregnancy and intrauterine fetal infection, microcephaly, brain damage, congenital malformation syndrome and evidences from laboratory models of fetal infections are reported [20–22]. The best prevention practice is the use of mosquito nets, protective clothing such as long-sleeve shirts and full pants and insect repellents to avoid the mosquito bites. In high endemic areas, it becomes important to use repellents on a routine basis to prevent bites from infected mosquitoes.

Several reports suggest that trace elements such as copper and cobalt play an important role in the host immunomodulatory activities during viral infections [23,24]. Any changes in the levels of trace elements could impact disease severity [25]. Essential oils from plants and herbs can inactivate enveloped and non-enveloped viral replication and exhibit antiviral activities [26–29]. Due to the strong antiviral and antimicrobial activities, novel citral-based compounds (the well-known natural insect repellent) are developed continuously [26–29]. Studies that show the effect of conjugation of citral-based compounds with different trace elements on ZIKV and dengue virus replication has not been explored. The purpose of the current study is to analyze the effects of copper(II) and cobalt(III) metal conjugations with organic compounds (that are used in repellents or food additives) on DENV2/ZIKV infections. The four compounds analyzed in this study are selected based on the structural activity relationship (SAR) in order to determine the best efficacy in inhibition of the virus. It has been reported that when “free” thiosemicarbazone ligands are coordinated to a metal center, the efficacy in the activity is enhanced in comparison to the “free” ligands. In this study, we show that treatment of mosquito (C6/36 cells) and human keratinocytes (HaCaT cells), HeLa cells and primary cultures of mouse bone marrow-derived dendritic cells with thiosemicarbazones coordinated to Cu(II) or Co(III) metal centers enhances ZIKV and DENV2 infections. Our data suggests that use of coordinated version with metal centers in the repellent formulations may aid better replication of ZIKV and DENV2 in the human cells upon transmission by the biting mosquitoes.

2. Materials and methods

2.1. Synthesis of the ligands and the compounds

Analytical or reagent grade chemicals were used throughout this study. All the chemicals including solvents were obtained from Sigma-Aldrich (St. Louis, MO, U.S.A.) or other commercial vendors, and used as received. Microanalyses (C, H, N) were performed by Intertek Chemical and Pharmaceuticals, 291 Route 22 East, Salem Industrial Park, Building #5, Whitehouse, NJ 08888, U.S.A. The FT IR (Fourier Transform infrared) spectrum was acquired in the range of 4000–400 cm−1 using the Attenuated Total Reflectance (ATR) accessory (with a diamond crystal) on a Nicolet 6700 FT IR spectrometer. 1H nuclear magnetic resonance (1H NMR) spectra were acquired on a Bruker AVANCE III 400 MHz spectrometer with deuterated dimethyl sulfoxide-d6 (DMSO-d6) as solvent. All 1H NMR spectra were processed with the Spectrus Processor 2012 software, which is available from Advanced Chemistry Development (ACD, Inc., 8 King Street East, Suite 107, Toronto, Ontario M5C 1B5, Canada).

The “free” salts and complexes

The “free” CuCl2·2H2O and CoCl2·6H2O salts were obtained from Sigma-Aldrich Corporation.

Synthesis of (E)-N-ethyl-2-[1-(thiazol-2-yl)ethylidene]hydrazinecarbothioamide (acetylatedTSC) 1 and [(Cu(acetylatedTSC)Cl)Cl0.25C2H5OH 2 (Compounds 1 and 2, respectively)

(E)-N-ethyl-2-[1-(thiazol-2-yl)ethylidene]hydrazinecarbothioamide (acetylatedTSC) 1 was synthesized and characterized as by Lewis et al.; [30] while the complex, [(Cu(acetylatedTSC)Cl)Cl0.25C2H5OH 2 (where
acetylthTSC = (E)-N-ethyl-2-[1-(thiazol-2-yl)ethylidene]hydrazinocarboxothioamide) was synthesized and characterized as by Sandhus et al. [31]

**Synthesis of citralEtTSC·0.25C2H5OH 3 (Compound 3)**

Citral (3.0 g, 3.377 ml, 20 mmol) and 4-ethyl-3-thiocarbazide (2.4 g, 20 mmol) were mixed in a 250 ml round bottom flask, followed by anhydrous ethanol (100 ml) and five drops of glacial acetic acid. The reaction mixture was refluxed with stirring for three hours and evaporated to dryness. The resulting oil was allowed to solidify at room temperature until it solidified to leave the resulting product. Yield = 4.8 g (91%). Calc. C13.6H24N4O6S6, C, 51.28; H, 2.34; N, 38.42%. Found: C, 51.28; H, 2.34; N, 38.42%. FT IR (ν/cm⁻¹): 3090 (C = N), (1296) (m) and (813) (m) (C = S). 1H NMR (400 MHz; CDCl3) δ: 3.28 (br, 2H, CH2), 7.31 (d, J = 8.8 Hz, CH), 10.78 (s, 1H, hydrazinic-NH), 13.2 (s, H-bonded hydrazinic-NH) ppm.

**Synthesis of [Co(phen)2(citralEtTSC)](NO3)3·0.25H2O·1.5C2H5OH 4 (Compound 4)**

At first, [Co(phen)2]Cl2·H2O was synthesized using the procedure as reported by Kashivbara et al. [32]. Then, the prepared [Co(phen)2]Cl2·H2O was synthesized using the acetylation of [Co(phen)2]Cl2·H2O by acetic anhydride. 1H NMR (400 MHz; DMSO-d6): δ_a = 5.0 (t, J = 10.3 Hz, CH2), 7.79 (d, J = 10 Hz, CH2), 7.99 (d, J = 10 Hz, CH2), 8.08 (d, J = 8 Hz, CH2), 8.30 (s, H, aminic NH), 10.58 (s, 1H, hydrazinic-NH), and 13.2 (s, H-bonded hydrazinic-NH) ppm.

2.3. RNA extraction, cDNA synthesis and Quantitative Real-Time PCR (Q-PCR) analysis

Total RNA from 1 × 10^5 of either C6/36 cells or HaCaT cells, first treated with the compounds (24 h) using different doses (1, 10 or 100 μM) followed by infection with DENV2/ZIKV (MOI 1; 24 h post infection) or post-treated (24 h) with respective compounds (indicated doses) after DENV2/ZIKV infection (MOI 1; 24 h) were generated using Aurum Total RNA Mini kit (BioRad, Hercules, CA) following manufacturer instructions. RNA was converted to cDNA using iScript cDNA synthesis kit (BioRad). The generated cDNA was used as template for the amplification of DENV2/ZIKV capsid/NS5 transcripts, respectively, from cDNA with primers as described [36,37]. As an internal control and to normalize the amount of template, either mosquito or mouse or human beta actin amplicons were quantified using published primers [36]. Q-PCR data showing DENV2/ZIKV loads were presented by normalizing expression of capsid/NS5 gene transcripts to either mosquito or mouse or human beta actin. Q-PCR was performed using iQ-SYBR Green Supermix and CFX96 touch System (BioRad). Standard curves were prepared using 10-fold serial dilutions starting from 1 to 0.00001 ng/μl of known quantities of DENV2/ZIKV, capsid/NS5, and respective beta-actin fragments. Following are the oligonucleotides used in this study: DENV capsid, 5′-ATATGCTGAAGACCGAGAGAGA- AACCCGCG 3′ and 5′-CTCTTCTATGTCAGTCCGAGTG 3′; ZIKV NSS, 5′-AARTAATCTACATACAAACAGTGTGTG 3′ and 5′-TCCRCCTCCCTYTG- GTCTG 3′; human actin, 5′-AGCTCGGCTGCGGCGCAGG 3′ and 5′-CTGG- TGGCTGGGCGG 3′; mouse actin, 5′-CTTACCAGGCAAGGTGTTGA 3′ and 5′-GGGTGTTGAGGTGGTCAACA 3′; mosquito actin, 5′-CCATG- TACGTGCCCTACCA 3′ and 5′-GGGTGTTGAGGTGGTCAACA 3′; human IFN-beta, 5′-CAGAAATTCTTTCAGTGAGAAGCT 3′ and 5′-TCATCTCTTCC- TTGAGGCAGT 3′; and human TNF-alpha, 5′-CCAGGGCAGTCAGATC- ATCTT 3′ and 5′-TCTCAGCTCCACGGCATT 3′.

2.4. Western blotting

Total proteins from 1 × 10^5 of either C6/36 cells or HaCaT cells infected with DENV2 (MOI 1; 24 h post infection) or with ZIKV (MOI 1 or 5; for 24 h) and then followed by treatments with respective compounds at different doses (1, 10, 100 μM) for another 24-h period or vice versa (treated with compounds and then infected; in a similar manner as described above) were collected in ice-cold modified RIPA buffer (G-Biosciences, BioExpress, Kaysville, UT). Total proteins were estimated using BCA kit. Western blotting, blocking (with either stain-free or BSA NAP-blocker solution; G-Biosciences, BioExpress) was processed and blots were incubated with 4G2 monoclonal antibody (non-reducing conditions), obtained from Dr. Michel Ledizet, L2 Diagnostics, Inc. Monoclonal Anti-Flavivirus Group Antigen, Clone D1-4G2-4-15 (Catalog Number; NR-50327), antibody obtained from BEI resources was used to detect ZIKV E-protein. Mouse monoclonal IgG conjugated with HRP (Santa Cruz Biotechnologies, Dallas, TX) were used as secondary antibodies for detection. Total protein profiles serve as loading controls and not for comparison between the compounds. The differences in protein profile gels are due to use of either stain-free gel images after run, or staining of gels with either colloidal Coomassie blue or R-250 Coomassie blue solutions. ECL detection of antibody binding was performed with WesternBright ECL kit (Advansta, BioExpress). Blots were scanned using Chemidoc MP imaging system and recommended Image Lab software from the manufacturer (BioRad).

2.5. Virus titration assays

For the titration assay, briefly, C6/36 or HaCaT cells were seeded
into 96 well plates and at a density of $1 \times 10^5$ cells/well in 225 μl of either MEM or DMEM complete medium with 10% FBS, pen/Strep and l-Glutamine. Cells were incubated overnight at either 30 °C or 37 °C with 5% CO2, respectively. Cells were treated with respective compounds 1–4 at doses of 1, 10 or 100 μM (for 24 h), followed by infection with DENV2 (MOI 1) for another 24 h. Each of the compound and respective doses were kept as six independent replicates in addition to the uninfected, infected and infected-DMSO treated controls. Following 24 h post-infection, cells were fixed with acetone-PBS mixture (3:1) for 20 min at −20 °C. Immediately after fixation, plates were air dried and washed with 1 × PBS and blocked with 5% FBS-PBS-0.05% Sodium Azide for 15 min at RT. After blocking, cells were incubated with 4G2 monoclonal antibody for overnight at 4 °C. Cells were washed three times with PBS and incubated with Alexa-594 labeled mouse secondary antibody for 1 h at RT, followed by washes (3 ×) with PBS. Plates were analyzed using EVOS fluorescence system (Invitrogen/Thermoscientific) and cells were scored for fluorescence or the presence or absence of infection in comparison to the positive (Infected and infected-DMSO-treated) or negative (uninfected) controls. Percentages were calculated from ratio of fluorescent cells to no fluorescent cells and as manual countings from eight microscopic fields. We collected eight images for each treatment and respective doses and representative images are shown in Supplementary information. Infection indicated by red fluorescence was determined as average from collected images and percentages were plotted in comparison to the infected-DMSO-treated controls.

### 2.6. Viral plaque formation assay

Plaque assays were performed as described [36]. For the plaque formation assay, briefly, Vero cells were seeded into 6 well plates and at a density of $1.5 \times 10^6$ cells/well in DMEM complete medium with 10% FBS. Cells were incubated overnight at 37 °C with 5% CO2 and then treated with compounds 1–4 at doses of 100 μM (for 2 h), followed by infection with DENV2 (MOI 1) for 2 h. Following post-infection, medium was removed and warm 2% Seaplaque agarose (Lonza) overlay with complete DMEM media (1:1 ratio) containing antibiotic and antimycotics solution (1% each; Sigma) was added. Plates were incubated for additional 5 days, at 37 °C and 5% CO2. After an incubation period, plaques were stained with 0.03% of Neutral Red (Sigma) for 4 h; stain was removed to count plaques. Each of the compounds (at the dose of 100 μM) was kept as two independent replicates in addition to the controls (infected and infected-DMSO treated). Representative images are shown for each compound and controls. Plaques counting are averages from three independent readouts.

### 2.7. Isolation and culturing of mouse bone marrow-derived dendritic Cells

Bone marrow cells were isolated from femurs and tibias of C57BL/6 mice, and red blood cells were lysed with ammonium chloride. Remaining cells were plated at a density of $7 \times 10^6$ per 100-mm cell culture dish in 10 ml of α-MEM media (HyClone) supplemented with 10% Fetal Bovine Serum (Atlanta Biologicals), 2 mM l-glutamine, 10% FBS. Cells were incubated overnight at 37 °C with 5% CO2 and then treated with respective doses of 1, 10 or 100 μM (for 24 h), followed by infection with DENV2 (MOI 1) for another 24 h. Each of the compound and respective doses were kept as six independent replicates in addition to the uninfected, infected and infected-DMSO treated controls. Following 24 h post-infection, cells were fixed with acetone-PBS mixture (3:1) for 20 min at −20 °C. Immediately after fixation, plates were air dried and washed with 1 × PBS and blocked with 5% FBS-PBS-0.05% Sodium Azide for 15 min at RT. After blocking, cells were incubated with 4G2 monoclonal antibody for overnight at 4 °C. Cells were washed three times with PBS and incubated with Alexa-594 labeled mouse secondary antibody for 1 h at RT, followed by washes (3 ×) with PBS. Plates were analyzed using EVOS fluorescence system (Invitrogen/Thermoscientific) and cells were scored for fluorescence or the presence or absence of infection in comparison to the positive (Infected and infected-DMSO-treated) or negative (uninfected) controls. Percentages were calculated from ratio of fluorescent cells to no fluorescent cells and as manual countings from eight microscopic fields. We collected eight images for each treatment and respective doses and representative images are shown in Supplementary information. Infection indicated by red fluorescence was determined as average from collected images and percentages were plotted in comparison to the infected-DMSO-treated controls.

**Fig. 1.** Treatment with organic compounds post infection enhances dengue virus loads. Q-PCR and immunoblotting analysis showing the DENV2 capsid transcripts or envelope E-protein loads in C6/36 mosquito cells (A & B) or human keratinocytes (HaCaT cells) (C & D) infected with DENV2 (MOI = 1) for 24 h followed by treatment with compounds 1–4 at different concentrations (1, 10 and 100 μM) for another 24 h, respectively. DENV2 capsid mRNA transcript levels (A & C) were determined by Q-PCR upon treatment with compounds in comparison to the infected-DMSO controls. UT indicates untreated cells; M shows marker lane and μM means micro molar. Uninfected (UI) or infected (I) cells treated with DMSO are included as controls. DENV2 E-protein was detected in panels B & D and upper band indicates glycosylated form. Total protein profiles are shown as loading control and not for comparison. The differences in protein profiles are due to use of Colloidal Coomassie blue staining in panels with compounds 1 and 4 and Coomassie blue R-250 staining solution in panels with compounds 2 and 3. Statistics was performed using the Student's two-tailed test. The $P < 0.05$ is indicated with an asterisk to show significance in comparison to DMSO-treated control and error bars indicate standard deviation. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)
dextrose, essential and non-essential amino acids, sodium pyruvate, sodium bicarbonate, antibiotics, and 2-β-mercaptoethanol. Granulocyte-macrophage colony-stimulating factor (GM-CSF; Peprotech; 5 ng/ml) was added at the time of plating, and again after 72 h of incubation. Experiments using animals were in accordance with protocols from the Institutional Animal Care and Usage Committee (as per the approved protocol 14–009). For infection and treatment with compounds, we re-plated dendritic cells into 24 well plates as 1 × 10⁵ cells/ml in each well and infected with either DENV2/ZIKV (MOI = 1), respectively. After 24 h post infection, dendritic cells were treated with all four compounds at different doses (1, 10, 100 μM) for another 24 h, and collected for RNA extractions.

2.8. Statistics

Statistical significance of differences observed in data sets was analyzed using Prism6 software (GraphPad, La Jolla, CA) and Microsoft Excel. For data to compare two means, the non-paired, two-tail Student t-test was performed for all analysis. P values of < 0.05 were considered significant in all tests, error bars represent mean (+SD) values. Statistical test and P values are shown.

3. Results

3.1. Synthesis and schemes of compounds

(E)-N-ethyl-2-[1-(thiazol-2-yl) ethylidene]hydrazinecarbothioamide (acetylethTSC) compound 1 was synthesized as described by Lewis et al.;[30] while the compound 2, [Cu(acetylethTSC)Cl]0.25C2H5OH 2 (where acetylethTSC=(E)-N-ethyl-2-[1-(thiazol-2-yl)ethylidene]hydrazinecarbothioamide) was synthesized as described by Sandhaus et al.[31] (Scheme 1, Supplementary Information). Compounds 3 and 4 were synthesized as described in (Scheme 2, Supplementary Information). The 1H NMR spectrum of compound 3 (citralEtTSC·0.25C2H5OH) is shown in Supplementary Fig. 1A. The thiosemicarbazone ligand derived from 2-acetylethiazole (compound 1), 2-acetylthiazole thiosemicarbazone ligand-copper(II) complex (compound 2), had been synthesized in [31]. Compounds 3 and 4 that are thiosemicarbazone ligands derived from citral without (compound 2) or with Co(III) complex (compound 4), respectively, were synthesized in this study as shown in Schemes 1 and 2. Complexes 5 and 6, [Cu(phen) 2]Cl2 and [Co(phen)3]Cl3, respectively, were synthesized in this study as by the literature procedure [34,35]. Schemes 3 and 4 in Supplementary information show the synthesis of complexes 5 and 6. The 1H NMR
spectrum of complexes 5 and 6 and [Co(phen)₃]Cl₂ are shown in Supplementary Fig. 1B and 1C. The methods used to synthesize these compounds are validated and are routinely been used in the biochemistry laboratory [31].

3.2. Thiosemicarbazone and 1, 10-Phenanthroline ligands conjugated with Cu(II) or Co(III) complexes enhance dengue virus loads

We determined the dengue virus in order to investigate any interference of these compounds in virus inhibition. C6/36 cells or HaCaT cells were infected with DENV2 (MOI = 1) for 24 h, followed by 24-h of incubations with respective compounds at different doses of 1, 10 and 100 μM concentrations. Q-PCR data showed that treatment of C6/36 cells with compounds 1 and 3 (100 μM) and compounds 2 and 4 (10 and 100 μM) concentrations significantly (P < 0.05) enhanced the DENV2 capsid mRNA transcripts in comparison to the infected-DMSO control (Fig. 1A). Immunoblotting analysis showed increased dengue envelope (E) protein loads in C6/36 cells treated with compound 2 or 4 at all concentrations (1, 10 and 100 μM) in comparison to DMSO-treated control (Fig. 1B). Human keratinocytes treated with compounds 2 and 4 (10 and 100 μM) also showed significant (P < 0.05) increase in dengue capsid mRNA transcripts in comparison to the infected-DMSO-treated control (Fig. 1C). However, compound 1 showed significantly increased dengue capsid mRNA transcripts at 100 μM treatment (Fig. 1C). Immunoblotting analysis with HaCaT cells revealed that compounds 2 and 4 (at 10 and 100 μM) showed dramatic increase in dengue E-protein levels in comparison to the infected-DMSO treated control (Fig. 1D). Increased dengue virus E-protein loads were evident in compound 1-treated cells even at 100 μM concentrations in comparison to infected-DMSO-treated controls. Total protein profiles serve as loading control and are not shown for comparison to the infected (untreated) or infected-DMSO treated samples (Fig. 1B & D). In addition to compounds 2 and 4, we tested complexes 5 and 6 to find if coordination of copper(II) or cobalt(III) to these 1, 10-phenanthroline compounds also enhances DENV2 upon treatment. We found that in both mosquito and HaCaT cells, complexes 5 and 6 also enhanced DENV2 loads upon 24-h treatment followed by 24 h post-infection (Supplementary Fig. 2A and 2B). Viral loads were enhanced in a dose dependent manner when complexes 5 and 6 were treated at increasing concentrations of 1, 10, 100 μM (Supplementary Fig. 2A and 2B). In order to include the possibility that the inorganic salts containing Cu(II) or Co(II) may itself enhance the DENV2 loads, we tested the CuCl₂·2H₂O and CoCl₂·6H₂O salts independently in both mosquito and HaCaT cells. We found that in both cell lines the salts alone did not enhanced the DENV2 loads at the similar treatments (with tested doses of 1, 10, 100 μM) and infection times (Supplementary Fig. 2C and 2D). This data suggested that Cu(II) or Co(II) salts alone has no effects as free cations.

3.3. Cu(II) or Co(III) metal centers with thiosemicarbazone ligands enhance production of infectious viral particles

To further confirm the enhanced viral replication and increased DENV2 loads upon treatment with compounds (2 and 4), we performed the virus titration and plaque formation assays as described in methods. In both mosquito and HaCaT cells, virus titration assay showed percentage of infected cells (determined by fluorescent signals). Both compounds 2 and 4, showed significantly (P < 0.05) increased percentage of infected cells at all tested doses of 1, 10 and 100 μM, for 24 h of treatment followed by another 24 h post-infection period (Fig. 2A and B). All four compounds (1–4) at 100 μM treatment showed increased production of viral infectious particles in both mosquito and human cells (Fig. 2A and B). However, compounds 2, 3 and 4 showed increased virus loads even at lower concentration of 10 μM in both cells (Fig. 2A and B). Representative images are shown for increased number of infected cells with pre-treatment of compounds 1–4 in both mosquito and HaCaT cells in comparison to their respective infected-DMSO treated controls (Supplementary Figs. 3 and 4). The plaque formation assay further confirmed the increased production of viral plaques in Vero cells treated with compounds 1–4 at 100 μM (Fig. 2C and D). The plaques formed in the presence of compounds 3 and 4 were large with bigger diameter in comparison to the plaques formed in infected-DMSO-treated controls (Fig. 2C). Also at 100 μM treatments, compound 2 showed several but smaller plaques in comparison to compound 1. The numbers of plaques formed were significantly increased upon treatment with compounds 2 and 4 in comparison to the compounds 1 and 3 (Fig. 2D). This data further confirmed that treatment with compounds containing Cu(II) or Co(III) metal centers increased production of infectious viral particles. Also, we analyzed the antiviral/cytokine response in DENV2-infected HaCaT cells treated with different compounds. Treatment of HaCaT cells with compound 1 (10 μM), showed significantly decreased levels of IFN-beta transcripts in comparison to infected-DMSO-treated control (Supplementary Fig. 5A). No differences were evident with the treatment with other compounds. In addition, we

Fig. 3. Treatment of compounds enhances DENV2 and ZIKV loads in primary cultures of mouse bone marrow derived dendritic cells. Q-PCR analysis showing DENV2 loads in primary cultures of mouse bone marrow derived dendritic cells showing DENV2 (A) and ZIKV loads (B). 1 × 10⁵ Cells were infected with MOI = 1 of either DENV2 or ZIKV for 24 h and followed by treatments with compounds 1–4 at different concentrations (1, 10 and 100 μM) for another 24 h, respectively. DENV2 capsid mRNA transcript (A) and ZIKV NS5 mRNA transcript levels (B) were determined by Q-PCR upon treatment with compounds in comparison to the infected-DMSO controls. UT indicates untreated cells and μM means micro molar. Uninfected (UI) or infected (I) cells treated with DMSO are included as controls. ns indicates not significant. Statistics was performed using the Student’s two-tailed test. The P < 0.05 is indicated with an asterisk to show significance in comparison to DMSO-treated control and error bars indicate standard deviation.
noted that treatment of HaCaT cells with compounds (1, 2 and 4) at 1 μM concentration showed significantly ($P < 0.05$) decreased levels of TNF-alpha transcripts (Supplementary Fig. 5B). Due to the observation of increased viral replication upon treatment with compound 4, we determined whether treatment with this compound elicits similar effect in additional human cell lines such as HeLa cells. QPCR analysis revealed that HeLa cells treatment with compound 4 at 100 μM showed significantly ($P < 0.05$) increased DENV2 loads, in comparison to the infected DMSO-controls (Supplementary Fig. 5C).

3.4. Thiosemicarbazone ligands conjugated with Cu(II) or Co(III) enhance DENV2 and ZIKV loads in primary cultures of marine bone marrow derived dendritic cells

We next determined DENV2/ZIKV loads in the primary cultures of mouse dendritic cells isolated from bone marrow derived precursor cells. Dendritic cells were infected with either DENV2 or ZIKV (MOI = 1) for 24 h, and then treated with compounds 1–4 at different doses (1, 10 and 100 μM) for another 24 h. QPCR analysis showed that both DENV2 (Fig. 3A) capsid transcripts and ZIKV NS5 transcripts (Fig. 3B) were significantly ($P < 0.05$) increased in Cu(II) conjugated compound 2 (both 10 and 100 μM treatments), however, unconjugated compound 1 showed significant ($P < 0.05$) increase in only ZIKV loads. Compound 3 treatments (at 100 μM) showed enhanced viral loads in both DENV2 and ZIKV infected dendritic cells (Fig. 3A & B). In dendritic cells infected with DENV2, treatment with compound 4 showed a significant ($P < 0.05$) increase in virus loads. However, treatments with compound 4 (100 μM) showed enhanced DENV2/ZIKV loads in comparison to their respective controls (Fig. 3A & B).

3.5. Treatment with copper(II) conjugated compound post infection with ZIKV enhances viral burden in both mosquito and human cells

In C6/36 cells infected with ZIKV (MOI = 1; for 24 h), followed by treatment with compounds 1–4 (at 1, 10 and 100 μM) for another 24 h, we found that ZIKV loads (NS5 mRNA transcripts) were significantly ($P < 0.05$) enhanced upon treatment with compound 2 (at 100 μM) (Fig. 4A). Although, no detectable levels of E-protein were evident in the same samples at MOI = 1 infectious dose (Fig. 4B). It was noted that in HaCaT cells, under similar conditions (ZIKV infection, MOI = 1, for 24 h; followed by compounds (1–4) treatments at 1, 10 and 100 μM for another 24 h), ZIKV NS5 mRNA transcripts were significantly ($P < 0.05$) increased upon treatment with compound 2, at both 10 and 100 μM concentrations (Fig. 4C). Also, ZIKV E-protein loads were elevated upon treatment with compound 2 (at 100 μM) in comparison to the infected DMSO-treated controls (Fig. 4D). Other compounds and their tested doses did not show any significant changes at either mRNA or protein levels (Fig. 4). We did not detect any ZIKV E-protein in infected and infected DMSO-treated controls due to very low infection with MOI = 1. Total protein profiles serve as loading controls (Fig. 4B & D). These results suggest that treatment with Cu(II) conjugated compound 2 post infection with ZIKV enhance viral burden, whereas without conjugation the same compound 1 did not showed any enhancement in ZIKV loads.

Since we did not find any detectable levels of E-protein in infected (untreated) or infected DMSO-treated controls at MOI = 1 of ZIKV infection, we performed additional experiments with higher infectious doses of MOI = 5. We first infected C6/36 cells or HaCaT cells with ZIKV at higher multiplication of infection of 5 for 24 h, followed by

![Fig. 4](image-url)
treatments with compounds (1–4) at different doses of 1, 10, and 100 μM concentrations. We detected E-protein in all compound treatments including compound 2, and there were no differences in comparison to the infected DMSO-treated controls (Fig. 5A). When HaCaT cells were infected with MOI = 5 of ZIKV in a similar manner, we detected E-protein in all samples including the infected (untreated) and infected DMSO-treated controls (Fig. 5B). We also included sodium salicylate (NaSal) compound that inhibits flavivirus replication [38] as control. NaSal-treated (5 mM) cells showed inhibition of ZIKV replication and lower E-protein loads in both C6/36 cells and HaCaT cells (Fig. 5A & 5B). Total protein profiles serve as loading controls in both C6/36 cells and HaCaT cells (Fig. 5A & B).

3.6. Pre-treatment with compounds 2 or 4 before infection with ZIKV enhance viral burden in both mosquito and human cells

To analyze if treatment with compounds before infection also enhance ZIKV replication in C6/36 and HaCaT cells, we first treated cells with compounds 1–4 (at concentrations of 1, 10 and 100 μM) for 24 h and then infected with ZIKV (MOI = 1) for another 24 h. In C6/36 cells, treatment with Cu(II) or Co(III) conjugated- compounds 2 (10 and 100 μM) or 4 (100 μM) significantly (P < 0.05) increased ZIKV NSS transcripts in comparison to the infected (untreated) or infected DMSO-treated controls (Fig. 6A). It was also noted that ZIKV E-protein loads were enhanced in C6/36 cells treated with compound 2 (10 and 100 μM) or 4 (100 μM) (Fig. 6B). In HaCaT cells, inductions in ZIKV loads were also noted in treatments with both Cu(II) or Co(III) conjugated compounds 2 and 4 (100 μM), respectively at 100 μM concentration. In addition compound 1 (100 μM) also showed significant (P < 0.05) induction in ZIKV NSS loads in HaCaT cells (Fig. 6C). Immunoblotting showed that ZIKV E-protein loads were enhanced upon treatment with compound 2 (100 μM) in comparison to the infected DMSO-treated control (Fig. 6D). No significant changes were observed with other compounds and the tested doses (Fig. 6D). Total protein profiles serve as loading controls (Fig. 6C & D). Since E-protein levels were not detected in infected DMSO-treated or infected untreated controls, we next analyzed effect of these compounds on ZIKV infection with higher infectious doses of MOI = 5. We treated C6/36 cells and HaCaT cells with compounds (1–4) at different doses of 1, 10, and 100 μM concentrations for 24 h and then infected with ZIKV at higher infectious dose (MOI = 5) for another 24 h. With increased infectious dosage both C6/36 cells and HaCaT cells showed E-protein in all samples including the infected (untreated) and the infected DMSO-treated controls (Fig. 7). In C6/36 cells, no differences were noted between compounds 1–4, and in comparison to their respective infected DMSO-treated controls (Fig. 7A). However, a dose dependent enhancement of ZIKV loads was evident in HaCaT cells upon treatment with compounds 1–4 (Fig. 7B). Sodium salicylate treated (5 mM) C6/36 cells and HaCaT cells serve as controls and show inhibition of ZIKV replication and lower E-protein levels (Fig. 7A & B). Total protein profiles serve as loading controls in both C6/36 cells and HaCaT cells (Fig. 7A & B). Collectively, these results suggest that Cu(II) or Co(III) conjugated compounds, enhance ZIKV and DENV2 infection in mosquito cells and human keratinocytes.

4. Discussion

Mosquito-borne diseases such as malaria, filariasis, arbovirus encephalitis and dengue are some of the major human health problems in tropical and subtropical countries. Recently, there has been an outbreak of ZIKV and Yellow Fever virus infections in the Americas. One of the potential interventions to combat mosquito-borne diseases is the continuous use of repellents in children and adults [7]. Our study suggests that use of Cu(II) or Co(III) containing metal centers in future preparations of food additives/repellents against mosquitoes may enhance replication and production of infectious viral particles of both DENV2 and ZIKV in the human cells upon transmission by the biting arthropods. Use of some insect repellents has been discouraged due to their adverse effects [3–8]. Safe use of alternatives like repellents of natural origin such as essential oils and their derived complex products has been preferred. Several studies have shown that plant based essential oils such as lemongrass; allspice oils, citral and other compounds are effective in controlling insect nuisance, mosquito bites and exhibit repellling properties [1,29,39–41]. Repellent bioassays have revealed that the activity of lemon essential oils is due to the presence of citral component [42,43]. Our study has shown that thiosemicarbazone ligands derived from citral and 2-acetylethiazole and their conjugation with Co(III) or Cu(II), enhance ZIKV and DENV2 loads at both mRNA and protein levels, respectively. Testing of additional complexes 5 and 6 with Cu(II) and Co(II) salts alone did not show any significant increase in DENV2 loads, suggesting no effects of free cations on viral replication. Our data also suggested that conjugation with these metal complexes might affect the antiviral or antimicrobial properties of such compounds. The reduced loads of IFN-beta suggested a diminished antiviral response in human cells (Supplementary Fig. 5A).

Several studies have shown that plant compounds have antiviral activities against dengue virus and other enveloped viruses [44–47]. However, conjugation of Co(III) or Cu(II) to such compounds may increase DENV2 loads in mosquito and HaCaT cells suggesting that both arthropod and human cells are susceptible to metal centers that may interfere and enhance DENV2 replication. In addition to human keratinocytes, human epithelial cells (HeLa cells) showed similar increase in DENV2 replication at 100 μM doses, suggesting similar activity of the compound 4 in enhancing viral loads (Supplementary Fig. 5C). Primary cultures of mouse bone marrow derived dendritic cells infected with DENV2/ZIKV also suggested that compounds 2 and 4 increased viral replication at 100 μM concentration in other mammalian cells.
Infection with ZIKV, followed by treatment with compounds also showed similar results with compound 2, where 10 and 100 μM induced the ZIKV loads in both C6/36 cells and HaCaT cells (Fig. 4A & C). Although, upon treatment with compound 2, the ZIKV E-protein loads were higher in HaCaT cells (Fig. 4C & D), suggesting high susceptibility and less tolerance in presence of ZIKV with treatment of compounds conjugated to Cu(II) containing complexes. Increased infection with ZIKV (MOI = 5) showed that the E-protein loads in both mosquito and human keratinocytes had no changes in comparison to the infected-DMSO controls. UT indicates untreated cells and ns indicate not significant. Uninfected (UI) or infected (I) cells treated with DMSO are included as controls. M shows marker lane. Total protein profiles are not shown for comparison and serve as loading controls. Statistics was performed using the Student’s two-tailed test. The P < 0.05 is indicated with an asterisk to show significance in comparison to DMSO-treated control and error bars indicate standard deviation.

(Fig. 3B & C). Infection with ZIKV, followed by treatment with compounds also showed similar results with compound 2, where 10 and 100 μM induced the ZIKV loads in both C6/36 cells and HaCaT cells (Fig. 4A & C). Although, upon treatment with compound 2, the ZIKV E-protein loads were higher in HaCaT cells (Fig. 4C & D), suggesting high susceptibility and less tolerance in presence of ZIKV with treatment of compounds conjugated to Cu(II) containing complexes. Increased infection with ZIKV (MOI = 5) showed that the E-protein loads in both mosquito and human keratinocytes had no changes in comparison to the loads of E-protein from the infected (untreated) or infected DMSO-treated controls. This data revealed that with higher infectious dose of MOI = 5, and post-treatment of compounds after infection may moderately allow compounds to interfere in the enhancement of viral replication.

Pre-treatments with compounds 2 and 4 followed by ZIKV infection showed increased viral loads (NS5 mRNA transcripts) in both mosquito and human keratinocytes (Fig. 6A & C). However, the observation of higher loads of ZIKV E-protein in mosquito cells indicated greater replication of ZIKV in mosquito cells at all tested doses of compounds 2 and 4 (Fig. 6B). It was also noted that ZIKV loads in HaCaT cells (that were first treated with compounds 1, 2 and 4, followed by infection) were much higher when compared to the viral loads in HaCaT cells that were exposed to compounds after virus infection (Figs. 4C & 6C). It is assumed that when these Cu(II) or Co(III) conjugated compounds 2 & 4 are applied to normal human skin, there are greater chances of encountering enhancement in the ZIKV replication upon bites from infected mosquitoes. It was noted that HaCaT cells treated with compounds 1–4 for 24 h, then followed by ZIKV infection at higher infectious doses (MOI = 5), showed a dose dependent increase of ZIKV E-protein levels with all tested compounds and their respective increasing doses (Fig. 7B). This data suggested that pre-exposure of human keratinocytes to these conjugated compounds might better enhance ZIKV loads when confronted through the infected mosquito bite. It is also intriguing to note that treatment with compound 2, followed by infection (with higher infectious dose of MOI = 5) with ZIKV, dramatically increased the viral loads in HaCaT cells (Fig. 7B), suggesting conjugation of Cu(II) or Co(III) complexes interference with ZIKV replication. Irrespective of the treatment procedures (infection followed by treatment with these compounds or pretreatment of compounds followed by infection) incubations lead to enhancement in DENV2/ZIKV loads in both arthropod and human cells. This enhancement of DENV2/ZIKV infection suggest that Cu(II) or Co(III) complexes in food additives or repellent sprays, lotions or any other forms might be disadvantageous on human cells and perhaps on other animals and aquatic environments. The mechanism of action of these compounds in enhancing DENV2/ZIKV loads in human keratinocytes is a future perspective. In summary, the present study shows that pre/post treatment with organic compounds containing Cu(II) or Co(III) metal complexes enhances ZIKV/DENV2 infection in mosquito, mouse and human cells.

Transparency document

The Transparency document associated with this article can be found in online version.
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Author contributions

SD, SK and HS performed experiments. MJC, AH, JFA (Jessa), AM and AAH prepared compounds tested in this study. CS and PJK provided bone marrow derived dendritic cells used in this study. SD, GN, AAH, ML and JFA discussed, analyzed and interpreted the data. HS planned, designed, coordinated and compiled the entire study. LH, ML and JFA provided important reagents. All authors edited and approved the manuscript. HS supervised overall investigations, wrote the paper.

Appendix A. Supplementary data

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References